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(54) Title: DETECTION OF CRYPTOSPORIDIUM PARVUM		
(57) Abstract A method and kit for the detection of <i>Cryptosporidium parvum</i> in aquatic and biological samples such as surface water or feces and a vaccine and plasmid for inducing an immunologic response against <i>C. parvum</i> . The method relies on the use of primers to detect all or a portion of at least one DNA sequence characteristic of <i>C. parvum</i> , the sequence being all or part of the genomic regions referred to as 38G and HemA contained within recombinant plasmids pINV38G, and pHem4, respectively.		

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DETECTION OF CRYPTOSPORIDIUM PARVUM

The present invention relates to a method for the detection and/or identification of organisms and/or nucleic acid sequences of *Cryptosporidium parvum* (*C. parvum*) and to oligonucleotides and test kits containing them for use in said method and to plasmids, peptides and polypeptides for inducing an antibody response *in vivo* and to antibodies produced in response to such peptides, polypeptides and plasmids.

Cryptosporidium parvum (phylum Apicomplexa) is a coccidian protozoan capable of parasitizing the intestinal tract of a variety of mammalian species. First recognized as a veterinary pathogen, its importance as a cause of human disease was not appreciated until the development of the acquired immunodeficiency syndrome epidemic. *C. parvum* is now recognized as a significant cause of diarrheal disease in both immunosuppressed and immunocompetent people worldwide. The disease has an incubation period of 1-14 days with symptoms lasting for 7-60 days. This intestinal pathogen has also been shown to play a significant role in the vicious diarrhea/malnutrition cycle experienced by young children in developing countries. Numerous outbreaks of cryptosporidiosis have been reported in the United States associated with person-to-person contact in day-care centers and hospitals, and waterborne spread from contaminated public water supplies. Resistance to chlorination and lack of an effective treatment contribute to the organism's spread.

C. parvum exists in nature in the form of environmentally resistant, thick walled oocysts. Large numbers of oocysts have been recovered from waste surface and recreational waters, but their origin is unknown. These oocysts have been known to survive for considerable periods in water and are unaffected by conventional water treatments. After ingestion, motile sporozoites released from the oocysts by the action of bile acids within the gut lumen invade the epithelial cells lining the intestine, form parasitophorous vacuoles beneath the microvillus membranes of these host cells, and initiate a complex life cycle containing both sexual and asexual reproductive stages. *C. parvum* sporozoites contain an apical complex consisting of rhoptries, micronemes, and pre-conoidal rings which are believed to play a key role in host cell invasion. It has been postulated that proteins secreted from the apical complex of these life stages initiate alterations in enterocyte membranes that lead to successful intracellular localization of the parasite. Very little is known about the molecular mechanisms for adherence to and penetration of intestinal epithelial membranes by cryptosporidial sporozoite life stages. Identification of specific sporozoite invasion factors could provide sensitive targets for more effective treatment strategies. Unfortunately, the lack of efficient laboratory culture methods to obtain large quantities of *C. parvum* has hampered efforts to directly isolate and characterize cryptosporidial proteins expressed during host cell parasitization. Proteins involved in the invasive processes of other intracellular pathogens have been

indirectly identified by cloning genomic regions from these organisms into nonpathogenic *Escherichia coli* strains and screening the transformants for the ability to invade cultured cell lines and escape subsequent antibiotic exposure. The utilization of gentamicin survival assays to identify invasive proteins from human pathogens was first described in studies of *Yersinia pseudotuberculosis* invasion. Since that time, proteins involved in the invasive mechanisms of *Yersinia enterocolitica* and *Mycobacterium tuberculosis* have been identified in a similar manner.

DESCRIPTION OF THE INVENTION

ISOLATION OF THE 38G GENOMIC REGION

A *C. parvum* genomic expression library was transformed in *E. coli* XL1-Blue and the transformants were tested for invasive activity in gentamicin survival assays using an intestinal-like epithelial cell line. Described herein is the identification and characterization of a 1.3 kb *C. parvum* genomic region which induces invasive activity in a nonpathogenic *E. coli* strain.

Materials and Methods

Library formation. *C. parvum* oocysts were obtained from infected Holstein calves and purified as described by Kuhls, et al. (J. Protozool, 1991, 38:74S-76S). Approximately 2×10^9 oocysts were freeze-thawed eight times in liquid nitrogen and incubated overnight at 55°C in lysis buffer (120mM NaCl, 10 mM EDTA, 25mM

Tris Ph 7.5, 1% Sarkosyl) containing 1 mg/ml proteinase K. Nucleic acids isolated by phenol/chloroform extraction and ethanol precipitation were treated with Rnase to remove co-precipitating RNA. *EcoRI* linkers were ligated to randomly sheared chromosomal DNA and a library consisting of 6.3×10^6 primary recombinants was constructed in the *EcoRI* site of the Lambda ZAP II expression vector (Stratagene). A representative portion of the phage library was excised with R408 helper phage to generate subclones in the pBluescript SK-phagemid vector which were transformed in competent *E. coli* XL1-Blue cells.

Gentamicin Survival Assay. Intestine 407 (ATCC CLL-6) monolayers were maintained in RPMI-1640 medium with 5% fetal calf serum. The monolayers were incubated for 3 hours at 37°C with *E. coli* XL1-Blue library transformants at a ratio of 100 bacteria per cell. After incubation, the monolayers were washed several times with sterile phosphate buffered saline (Ph 7.4) and incubated in fresh media containing 75 ug/ml gentamicin for 1.5 hours at 37°C to destroy any remaining extracellular bacteria. The cells were lysed with 0.5% Triton X-100, and the cell lysates were plated on L agar containing 100 ug/ml ampicillin for identification and quantitation of surviving transformant strains. In the initial screening of the *C. parvum* genomic library, 1.6×10^5 *E. coli* XL1-Blue transformants were tested. Those strains surviving the first assay were scraped from the agar plates, reincubated with fresh tissue culture cells, and assayed again. Survivors from this second round were tested a third time. In assays testing

individual transformants, plate counts of the original inoculums were performed in order to determine a survival rate for each transformant. Survival rates were calculated as follows:

100 x [number of bacteria present in cell lysates/number of
5 bacteria added to monolayers].

Sequence analysis. Nucleotide sequences were determined utilizing ³²P end-labeled primers with the *fmol* DNA Sequencing System (Promega) according to the manufacturer's instructions. T3 and T7 sequencing primers were provided in the kit and additional
10 primers were synthesized by the Molecular Biology Resource Facility of St. Francis Hospital located at the University of Oklahoma Health Sciences Center. Homology searches and sequence analysis were performed using the University of Oklahoma Genetics Computer Group sequence analysis software package (version 6.0) from the
15 University of Wisconsin Biotechnology Center.

Analysis of plasmid encoded polypeptides. [³⁵S]methionine labeled plasmid encoded translational products were expressed in maxicells of *E. coli* CSR603 using the method described by Sancar, et al., (J. Mol. Biol., 1981, 148:45-62). Cell lysates were
20 separated by SDS-PAGE (Laemmli, U., Nature, 227:680-685) on a 16.5% gel, and proteins were visualized by exposure to Kodak X-Omat AR film.

Pulsed Field Gel Electrophoresis. Oocysts were prepared for electrophoresis by the method described by Mead et al.,
25 (J. Parasitol., 1988, 74:366-369. Briefly, 1x10⁸ purified oocysts were washed and resuspended in 50 µl of suspension buffer (75 Mm

NaPO₄, 65 Mm NaCl, 1% glucose), Ph 8.0. Then 50 µl of 1% low gelling temperature agarose (SeaPlaque GTG, FMC BioProducts, Rockland, Maine) previously melted in suspension buffer and cooled to 37°C was added and 100 µl aliquots of the mixture were placed
5 into molds on ice to gel. After solidification, each agarose block was incubated in 10 ml of lysis buffer (0.5 M EDTA, 1.0% sodium N-lauroylsarcosine, 2.0 mg/ml proteinase K), Ph 9.5, at 50°C for 48 hours. The blocks were then washed extensively and stored in 50 Mm EDTA, 10 Mm Tris Ph 8.0 at 4°C. Agarose blocks were cut in half
10 and loaded into the slots of a 0.7% agarose (Pulse Field Certified Agarose, BioRad) gel in 45 Mm Tris, 45 Mm boric acid, 1.25 Mm ethylenediamine tetraacetic acid buffer, Ph 8.0 (TBE). The DNA was fractionated on a CHEF-DR III pulsed field electrophoresis system (BioRad). Gels were run at 14°C for 40 hours with a voltage of
15 6v/cm, an angle of 120°, and switch time from 60-120 seconds. After depurination in 0.25M Hcl, the gels were washed, and blotted by capillary transfer to Zeta-Probe GT nylon membranes (Bio-Rad). The membranes were hybridized with a ³²P-labeled 1.3 kb fragment according to instructions supplied by the manufacturer and exposed
20 to Kodak X-Omat AR film.

Electron microscopy. INT 407 monolayers were incubated for varying times periods with the transformants as described above. After several washings with phosphate buffered saline, the monolayers were fixed in 2% paraformaldehyde-2.5% glutaraldehyde in
25 cacodylate buffer, and routinely processed for transmission and scanning electron microscopy.

RNA isolation and RT-PCR. *C. parvum* sporozoites were excysted from purified oocysts and prepared as previously described (Kuhls et al., J. Protozool, 38:74S-76S). The sporozoites were frozen overnight at -70°C in guanidinium isothiocyanate denaturing solution supplied in the RNA Isolation Kit (Stratagene, La Jolla, Calif.) and ground exhaustively in a cold mortar and pestle kept at -70°C. Total RNA was extracted according to the kit instructions. Isolated RNA was incubated at a concentration of 1 unit/ μ g of Rnase free Dnase (Stratagene) for one hour at 37°C to remove any contaminating DNA.

RT-PCR was performed on 650 ng of total sporozoite RNA per reaction with the reagents and instructions provided in the SUPERScript Preamplification System (Gibco/BRL, Gaithersburg, Md.). Random hexamers were used to prime first strand cDNA synthesis.

PCR amplifications were performed in a 100 μ l reaction mixture containing 50 pmoles of each primer and 5 units of Taq DNA polymerase (Gibco/BRL) per reaction with the following cycling parameters: initial denaturation at 94°C for 5 min, followed by 40 cycles of 94°C for 1 min, 50°C for 30 sec, 72°C for 3 min, and a final 8 min extension at 72°C. The reactions were performed in a Perkin-Elmer model 480 thermocycler (Perkin-Elmer Cetus, Norwalk, CT). Negative control reactions omitting the reverse transcriptase were run with each sample to ensure that the PCR products were not due to amplification of contaminating sporozoite DNA. Positive control reactions using primers specific to the *C. parvum* actin gene were performed to verify the integrity of the isolated RNA and

the suitability of the reaction conditions. Actin primers were designed based on the reported sequence of a *C. parvum* actin gene, GenBank accession number M69014. The forward primer covers positions 365-384 and the reverse primer covers positions 659-640 of the deposited nucleotide sequence. All oligonucleotides in this study were synthesized at the Molecular Biology Resource Facility, St. Francis Medical Research Institute, University of Oklahoma Health Science Center.

Results

Screening the bacterial transformants. Invasive activity was measured by testing the ability of the bacterial transformants to survive gentamicin exposure after incubation with Intestine 407 monolayers. Transformant strains closely associating with or penetrating the intestinal cell membranes were expected to be protected from the antibiotic. Seventy-six recombinant XL1-Blue transformants survived the three serial gentamicin survival assays. Recombinant plasmids isolated from sixty (79%) of these strains contained a 4.1 kb insert with identical restriction patterns demonstrated by agarose gel electrophoresis. Transformants containing this recombinant plasmid, designated pINV38G, were then tested individually in the survival assay along with strains containing nonrecombinant pBluescript and a recombinant pBluescript, pNHEM9, which was randomly selected from the plasmid library. pINV38G transformants consistently demonstrated a

capacity for gentamicin survival 500- to 4000-fold higher than pBluescript and pNHEM9 transformants (see Table 1).

TABLE I. Relative Survival of Transformants After Gentamicin Exposure of Intestine 407 Monolayers

INFECTING STRAIN	RELATIVE SURVIVAL ^a
XL1-Blue (pBluescript)	1
XL1-Blue (pINV38G)	500-4000
XL1-Blue (pINV38G1.3)	30-400
XL1-Blue (pNHEM9)	0.8-3

^aRelative survival values were determined by dividing the survival rate of the infecting strain by the survival rate of XL1-Blue (pBluescript). Survival rates = 100 x (# bacteria present after gentamicin exposure/# bacteria added to Intestine 407 monolayers). The ranges indicated reflect the results from several experiments.

Characterization of the cryptosporidial segment. Restriction mapping and nucleotide sequence analysis of the recombinant clone demonstrated that the 4.1 kb insert contained a 1.5 kb segment of pBluescript which was flanked on each side by identical 1.3 kb *C. parvum* genomic fragments. To establish that the survival phenotype of the pINV38G transformants was encoded on the 1.3 kb segment, gentamicin survival assays were performed with XL1-Blue cells freshly transformed with recombinant clones, designated pINV38G1.3, containing a single copy of the 1.3 kb region. These transformants demonstrated a lower level of gentamicin survival than XL1-Blue (pINV38G), but survival was consistently 30- to 400-

fold higher than that of pBluescript and pNHEM9 transformants (Table 1).

The nucleotide sequence of the 1.3 kb (38G) fragment had A+T content of 73%, consistent with that reported for other cryptosporidial genomic sequences. Computer analysis predicted several small open reading frames none of which appeared to have had significant homology to other reported sequences. Karyotypic Southern blot analysis of *C. parvum* chromosomal DNA fractionated by pulsed field gel electrophoresis mapped the 1.3 kb segment to the smallest of five fractionated *C. parvum* chromosomes. To identify protein products encoded by the 1.3 kb region, the recombinant plasmids were tested in maxicells. A radiolabeled protein of ca. 5,800 daltons was detected on autoradiographs of maxicell lysates from pINV38G and pINV38G1.3 transformants, but not in lysates from transformants containing pBluescript. The size of this protein is consistent with the size of two open reading frames predicted from the nucleotide sequence of the 1.3 kb fragment.

Electron microscopy of infected monolayers. To determine whether the bacterial transformants were actually internalized, both scanning and transmission electron microscopy were performed on Intestine 407 monolayers incubated from one to three hours with XL1-Blue(pINV38G) and XL1-Blue(pBluescript). The monolayers were washed extensively after the incubation periods, but gentamicin was omitted in order to ensure that adequate numbers of bacteria would be present for observation and comparison. After one and two hour incubations, most of pINV38G transformants were only loosely

associated with Intestine 407 cellular processes, however, some were more closely associated with the plasma membrane, and had one end turned toward the monolayers. After three hours, substantial numbers of bacteria had one end partially embedded into the intestinal cell membranes. The portion of each bacterium which was embedded ranged from <10% to 50%. Although portions of some bacteria were completely encircled by cell membrane and cytoplasm, the bacteria remained extracellular and were separated from the cytoplasm by plasma membrane in all areas observed. In contrast, although a similar number of bacteria were observed in all incubation periods with XL1-Blue(pBluescript), this nonrecombinant transformant remained only loosely associated with the Intestine 407 cells throughout three hours of incubation. In general, intestinal cell morphology was similar between the two samples studied, but monolayers exposed to pINV38G transformants had more cells which appeared damaged. These cells contained smoother surfaces (loss of cellular processes) and were more rounded than uninfected cells. pINV38G transformants were not seen embedded into the damaged cells.

Gentamicin survival in other tissue culture cell lines. To determine whether the invasive activity demonstrated by XL1-Blue(pINV38G) was specific to Intestine 407 monolayers, survival assays using other epithelial cell lines were performed. Compared to gentamicin survival present after incubation with Intestine 407 monolayers, XL1-Blue transformants demonstrated a five- to ten-fold higher level of survival in WISH cells, an equivalent level in Hep-

2 cells, a hundred-fold lower level in IEC-6 cells and no detectable survival in MDCK cells.

RT-PCR analysis of sporozoite RNA. Cryptosporidial genomic regions encoding proteins necessary for host cell entry should be actively transcribed in *C. parvum* sporozoites. To determine whether transcripts homologous to the 1.3 kb region were present in this life stage, total RNA isolated from *C. parvum* sporozoites was analyzed in RT-PCR reactions using a primer pair that was specific to an open reading frame in the 1.3 kb region. These primers are a forward primer shown in SEQ ID NO:5 and a reverse primer shown in SEQ ID NO:6. The anticipated amplification product of 152 bp expected from the primer pair was demonstrated on agarose gel electrophoresis of the RT-PCR reaction. The bands were excised from the gel and analyzed by nucleotide sequence analysis for verification of the PCR products.

Open Reading Frames

Bases 423 through 560 and bases 1134 through 1298 in SEQ ID NO:1. constitute a first and second open reading frame, respectively, in SEQ ID NO:1. Each of these open reading frames codes for a protein which can be used to detect the presence of *C. parvum*.

ISOLATION OF HEMA GENOMIC REGION

A *C. parvum* genomic expression library was transformed into a nonpathogenic *Escherichia coli* strain, and the transformants were screened on blood agar to identify hemolytic strains. Described
5 herein is the cloning and characterization of a cryptosporidial genomic region (HemA) which confers a hemolytic phenotype on *E. coli* XL1-Blue.

Materials and Methods

Library formation. Cryptosporidial oocysts were obtained from
10 experimentally infected Holstein calves and purified as previously described. Approximately 2×10^9 oocysts were freeze-thawed eight times with liquid nitrogen and incubated overnight at 55°C in lysis buffer (120mM NaCl, 10 Mm EDTA, 25mM Tris Ph 7.5, 1% Sarkosyl) containing 1 mg/ml proteinase K. Nucleic acids isolated by
15 phenol/chloroform extraction and ethanol precipitation were treated with Rnase to remove co-precipitating RNA. EcoRI linkers were ligated to randomly sheared chromosomal DNA and a library consisting of 6.3×10^6 primary recombinants was constructed in the EcoRI site of the Lambda ZAP II expression vector (Stratagene).
20 The phage library was excised with R408 helper phage to generate subclones in the pBluescript^{SK} phagemid vector.

Subcloning and sequence analysis. Unidirectional deletion clones were generated using exonuclease III (Stratagene) according to the manufacturer's instructions. Exonuclease III resistant
25 sites were created by NotI digestion of the recombinant plasmid

followed by end-filling with thioderivatives, and sensitive sites were created by digestion with *Bam*HI. Nucleotide sequences were determined utilizing ³²P end-labeled primers with the *fmol* DNA Sequencing System (Promega). The T3 and T7 sequencing primers were provided with the kit and all additional primers were synthesized by the Molecular Biology Resource Facility of St. Francis Hospital located at the University of Oklahoma Health Sciences Center.

Analysis of plasmid encoded polypeptides. Proteins encoded on H4 and the H4 deletion clones were identified by expression in maxicells of *E. coli* CSR603 by the procedure described by Sancar (*J. Bact.*, 1979, 137:692-693). Maxicell lysates were separated by SDS-PAGE on a 12.5% gel, and proteins were visualized by autoradiography on Kodak X-Omat AR film.

Pulsed Field Gel Electrophoresis. Cryptosporidial oocysts were prepared for electrophoresis by the method described by Mead (*J. Parasitol.*, 1988, 74:366-369). Briefly, 1x10⁸ purified oocysts were washed and resuspended in 50 µl of suspension buffer (75 mM NaPO₄, 65 mM NaCl, 1% glucose), pH 8.0. Fifty microliters of 1% low gelling temperature agarose (SeaPlaque GTG, FMC BioProducts, Rockland, Maine) previously melted in suspension buffer and cooled to 37°C was added and 100 µl aliquots of the mixture were placed into molds on ice to gel. After solidification, each agarose block was incubated in 10 ml of lysis buffer (0.5 M EDTA, 1.0% sodium N-lauroylsarcosine, 2.0 mg/ml proteinase K), pH 9.5, at 50°C for 48 hours. The blocks were then washed extensively and stored in 50 mM EDTA, 10 mM Tris pH 8.0 at 4°C. Agarose blocks were cut in half

and loaded into the slots of a 0.7% agarose (Pulse Field Certified Agarose, BioRad) gel in 22 mM Tris, 22 mM boric acid, 0.62 mM EDTA buffer, pH 8.0. The DNA was fractionated on a CHEF-DR III pulsed field electrophoresis system (BioRad). Gels were run at 14°C for 5 40 hours with a voltage of 6 v/cm, an angle of 120°, and switch time from 60-120 seconds. After depurination in 0.25M HCl, the gels were washed, and blotted by capillary transfer to Zeta-Probe GT nylon membranes (BioRad). The membranes were hybridized with a ³²P-labeled 5.5 kb fragment according to instructions supplied by the 10 manufacturer and exposed to Kodak X-Omat AR film.

Hemolytic assays. Overnight cultures of H4 and pBluescript XL1-Blue transformants were grown in 5 ml of terrific broth containing 100 µg/ml ampicillin. After centrifugation, the culture supernatants were kept on ice until tested. The cell pellets were 15 washed in sterile phosphate buffered saline (PBS), resuspended in 1 ml PBS, sonicated on ice and centrifuged to remove insoluble membranes. Sheep erythrocytes were washed in PBS and brought up to a 5% v/v suspension in PBS. For each assay, 500 µl of the test sample was incubated with an equal volume of 5% sheep erythrocyte 20 suspension for 4 hr at 37°C. After centrifugation, the release of free hemoglobin was measured by reading the optical density of the supernatant at 540 nm. The values for 0% and 100% erythrocyte lysis were determined by the addition of PBS and 0.1% Triton-X 100, respectively, to an equal volume of 5% sheep erythrocytes. 25 Sonicates were tested directly, after diluting 1:10 with PBS, in PBS containing 10 mM MgCl₂, or 10 mM EDTA, and in 10mM Tris-HCl

(pH7.2), 150 mM NaCl (TBS) containing 10mM CaCl₂. For assays in the presence of added Ca⁺⁺, the sheep erythrocytes were resuspended in TBS and sonicate dilutions were made with TBS. Sonicates were assayed after heat treatment at 60°C for 30 min and after
5 incubation with 200 µg proteinase K for 1 hr at 37°C. Protein concentrations in the sonicate supernatants were determined by the method used by Smith et al. (Anal. Biochem., 150:76-85, 1985).

RNA isolation. RNA was isolated from purified *C. parvum* sporozoites that were prepared as previously described (Kuhls et
10 al., J. Protozool., 1991, 38:74S-76S). The sporozoites were frozen overnight at -70°C in guanidinium isothiocyanate solution and then ground exhaustively with a cold mortar and pestle. Total RNA was isolated by phenol/chloroform extraction and incubated with RNase-free DNase (Stratagene) at a concentration of 1 unit/µg for one
15 hour at 37°C to remove contaminating DNA.

Male and female 6-week-old C.B-17Icr scid/scid (SCID) mice were given intragastric inoculations of 2x10⁶ purified cryptosporidial oocysts as previously described (Kuhls et al., J. Comp. Pathol., 1992, 106:399-410). Three uninfected and three
20 infected mice were killed by CO₂ inhalation at 60 days post inoculation and the ascending colons were removed, opened longitudinally and flushed to remove fecal debris. The ascending colons of one uninfected and infected SCID mouse were fixed in 10% buffered formalin for histopathologic analysis. The remaining
25 colonic segments were stored immediately at -70°C. The frozen intestinal tissues were ground exhaustively in guanidinium

isothiocyanate solution and total cellular RNA was isolated by phenol/chloroform extraction with incubation in RNase-free DNase as described above.

RT-PCR. For cDNA synthesis, 1 μ g of sporozoite RNA or 3 μ g of murine intestinal RNA was incubated for 1 hour at 42°C in a 20 μ l reaction mixture containing 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 3 mM Mg Cl₂, 0.4 μ g random hexamers (Gibco/BRL), 10 mM of each deoxynucleotide triphosphate, 20 units placental ribonuclease inhibitor (Gibco-BRL) and 200 units Moloney murine leukemia virus reverse transcriptase (Gibco-BRL). The samples were then heated at 94°C for 5 minutes, diluted to a final volume of 100 μ l with 0.1% (wt/vol) diethylpyrocarbonate treated water and stored at -20°C. PCR reactions were performed in a 50 μ l volume containing 15 μ l of the cDNA reaction, 10mM Tris-HCl (pH 8.3), 50mM KCl, 1.5 mM MgCl₂, 0.01% (wt/vol) gelatin, 50 pmoles of each primer, 10 mM of each deoxynucleotide triphosphate, and 5 units of Taq DNA polymerase. Twenty-five cycles consisting of 94°C for 1 min, 65°C for 2 min, 72°C for 1 min, and a final 8 min extension at 72°C were performed. Reaction products were visualized by electrophoresis of 35 μ l of the reactions in a 1.5 % agarose gel containing 1 μ g/ml ethidium bromide. The reactions were performed in a Perkin-Elmer model 480 thermocycler (Perkin-Elmer Cetus, Norwalk, CT). Negative control reactions which either omitted the reverse transcriptase or template were performed to ensure that the PCR products were not due to amplification of contaminating DNA. A forward primer for hemA is shown in SEQ ID NO:7. A reverse primer for HemA is shown

in SEQ ID NO:8. The cryptosporidial actin primers were designed based on the reported sequence of a *C. parvum* actin gene (Kim et al., Mol. Biochem. Parasitol, 1992, 50:105-114), GenBank accession number M69014. The forward primer (SEQ ID NO:7) covers positions 365-384 and the reverse primer (SEQ ID NO:8) covers positions 659-640 of the deposited nucleotide sequence SEQ ID NO:3. The oligonucleotide primers were synthesized at the Molecular Biology Resource Facility, St. Francis Medical Research Institute, University of Oklahoma Health Science Center. The mouse β -actin primers were the same as those used by Buchanan et al. (Infect. Immun., 62:2930-2939, 1994). They flank an intron in the mouse β -actin gene and amplify a 620-bp fragment from murine cDNA.

GenBank accession number. The nucleotide sequence for *hemA* was deposited in GenBank under accession number U18120.

Results

Identification of pHEM4. A cryptosporidial genomic expression library in the pBluescript phagemid was transformed into *E. coli* XL1-Blue and the transformants were plated on 5% sheep blood agar containing 100 μ g/ml ampicillin. After incubation at 37°C for 24 hours, recombinant strains with a hemolytic phenotype were distinguished by the development of a clear zone around the bacterial colonies. Those strains exhibiting hemolytic activity were restreaked for verification, and four stable hemolytic recombinant strains were identified. Dot blot hybridization studies and restriction mapping determined that two of these

strains contained a recombinant clone (designated H4) with the identical 5.5 kb cryptosporidial genomic insert. The H4 transformant exhibited the most striking hemolytic zone on blood agar plates compared to the other two hemolytic strains and was chosen for further analysis. To confirm that the hemolytic activity was encoded on the recombinant plasmid, fresh XL1-Blue cells transformed with H4, pBluescript and a nonhemolytic recombinant clone chosen randomly from the cryptosporidial genomic library were tested on blood agar. The new H4 transformants consistently caused hemolysis while transformants containing pBluescript and the nonhemolytic recombinant plasmid remained nonhemolytic.

Analysis of the H4 cryptosporidial insert. A set of overlapping deletion clones were generated from the 5' end of the 5.5 kb insert using exonuclease III. The subclones were transformed into XL1-Blue and the sizes of the remaining cryptosporidial inserts were determined. Transformants containing various lengths of the insert were plated on sheep blood agar to determine their hemolytic phenotype. Hemolytic activity was still detectable on blood agar after removal of 4.5 kb of the original insert. Nucleotide sequence analysis of the deletion clones demonstrated the presence of a complete 690 bp open reading frame (*hemA*) located within a 1 kb region at the 3' end of the original insert. XL1-Blue transformants containing subclones of the 5.5 kb insert which included only a portion of *hemA* did not demonstrate a hemolytic phenotype on sheep blood agar.

The nucleotide sequence of *hemA* is represented as SEQ ID NO:3 and its complementary sequence SEQ ID NO:4. The first ATG (position 71) in the open reading frame was identified as the putative initiation codon since there is a purine at position -3 consistent with other eukaryotic translation initiation signals and an in-frame stop codon that occurs immediately upstream. The high A+T content (69%) of *hemA* and the codon usage predicted from the primary amino acid sequence of the *hemA* gene product are similar to that described for other reported cryptosporidial genes. Computer analysis of the nucleotide sequence using GRAIL (Guan et al., Proc., The Eight IEEE Conference on AI Applications, 1992, pp. 9-13) and the University of Wisconsin Genetics Computer Group sequence analysis package version 7.0, (Devereux et al., Nucleic Acids Res., 1984, 12:387-395), predicted *hemA* to be a likely protein coding region. There were no identifiable polyadenylation signals in the remaining 180 bp of H4 insert downstream of the *hemA* stop codon. The 229-amino acid protein predicted by the *hemA* nucleotide sequence is predominantly hydrophilic and has a calculated molecular mass of about 26 kD. No membrane spanning regions or signal sequences are evident from the deduced primary amino acid sequence. Asparagine residues make up 11% of the predicted polypeptide. One potential N-glycosylation site is present, and there are numerous serine and threonine residues that could serve as sites of O-linked glycosylation. No conserved peptide motifs were found, but the tetrapeptide IENG is repeated four consecutive times within the carboxy terminus of the predicted

protein. A BLAST search of the current DNA and protein databanks revealed no significant homologies to *hemA* or its predicted protein.

Karyotypic Southern blot analysis. To confirm that the 5.5 kb insert of H4 was cryptosporidial in origin, chromosomal DNA isolated from *C. parvum* oocysts was fractionated by pulsed field gel electrophoresis, capillary blotted to a nylon membrane and hybridized with a ³²P-labeled 5.5 kb fragment. Five distinct chromosomal DNA bands were identified on ethidium bromide stained agarose gels. Hybridization studies mapped the cryptosporidial genomic region to the second smallest chromosome.

In vitro expression of the cryptosporidial protein. [³⁵S]methionine labeled plasmid-encoded proteins of H4 and the H4 deletion clones were expressed in maxicells of *E. coli* CSR603 to identify the cryptosporidial proteins encoded on the recombinant plasmids. A protein of about 22 kDa was apparent on autoradiographs of maxicell lysates from strains transformed with H4 and H4 deletion clones containing the complete *hemA* coding region, but not in lysates from transformants containing nonrecombinant pBluescript or a H4 deletion clone containing only a portion of *hemA*. The size of this protein is slightly smaller than the calculated molecular weight based on the predicted amino acid composition of the *hemA* encoded protein.

Hemolytic activity of H4. To further characterize the hemolytic activity of the H4 transformants, hemolytic assays were performed on culture supernatants and cell sonicates of H4 and

pBluescript transformants. The H4 culture supernatants did not exhibit significant hemolytic activity when added to sheep erythrocytes, indicating that the hemolytic activity encoded on H4 was not actively secreted into the culture media. However, 5 sonicates of H4 transformed cells displayed strong hemolytic activity when compared to sonicates of pBluescript transformed cells and to a 0.1% Triton-X 100 positive control (Table II). Incubation of erythrocytes with undiluted H4 cell sonicates resulted in almost complete erythrocyte lysis after four hours of 10 incubation. Dilution of the H4 sonicates 1:10 with PBS prior to incubation resulted in about 50% lysis of the erythrocytes. Proteinase K digestion and heat treatment of the H4 sonicates prior to the assay markedly decreased their hemolytic activity, while the addition of Ca^{++} , Mg^{++} and EDTA had no effect on the assay results. 15 Storage of the undiluted cell sonicates at 4°C and -20°C for one week did not appreciably diminish the hemolytic activity. Undiluted sonicates assayed at different incubation temperatures, displayed similar hemolytic activity at room temperature that was only mildly decreased when incubated at 4°C or on ice.

TABLE II. Location and Measurement of Hemolytic Activity In Overnight Cultures of H4 Transformants

% Hemolysis ^a		
	pBluescript	H4
5 Culture supernatants	4	7
Cell sonicates ^d		
Undiluted	14 ^b	94 ^b
Diluted ^c	1	52

10 ^a Hemolytic activity is expressed as a percentage of sheep erythrocytes lysed after 4 hours at 37°C. Percent hemolysis is calculated based on Triton-X 100 control for 100% lysis. All assays were performed in duplicate.

^b Protein concentration 2.7 mg/ml

^c Diluted 1:10 in phosphate-buffered saline (PBS)

15 ^d Cell pellets were resuspended in 1 ml PBS and sonicated on ice in multiple 10 second bursts with a Branson ultrasonifier.

Transcriptional analysis of *hemA*. Since membrane disruption is an integral part of the cryptosporidial life cycle, it is reasonable to expect that membranolytic proteins are made in many, if not all, of the protozoan's life stages. In order to determine whether *hemA* is actively transcribed during the sporozoite life stage, RT-PCR analysis was performed on total RNA isolated from excysted sporozoites using primers designed to amplify a 369 bp fragment internal to *hemA*. Positive control reaction were performed using primers designed to amplify a 295 bp region within a *C. parvum* actin gene (Kim et al., Mol. Biochem. Parasitol, 1992, 25 50:105-114). Agarose gel electrophoresis of the RT-PCR reactions yielded the expected gene fragments for each primer set. All

amplification products were verified by Southern blot hybridization studies using radiolabeled probes that had been previously confirmed by nucleotide sequence analysis to match the expected regions of *hema* or the cryptosporidial actin gene. Amplification with the *hema* primers gave an additional band of 270 bp that did not hybridize to the *hema* probe. Negative control reactions which omitted either the reverse transcriptase or template did not yield a PCR product.

Adult SCID mice experimentally infected with cryptosporidial oocysts are known to contain numerous organisms in the villous and crypt mucosa of their anterior colons by 60 days post inoculation. In order to determine whether *hema* is actively transcribed by *C. parvum* during infection of mammalian hosts, RT-PCR analysis of total RNA isolated from the ascending colons of infected mice was also performed. RNA isolated from the proximal colons of age-matched uninfected mice was used in control reactions. A 369 bp amplification product was present on agarose gels of reactions performed with RNA isolated from the intestines of infected mice but not with RNA isolated from uninfected mice. Mouse β -actin primers were included in the RT-PCR reactions as a positive control to verify that intact RNA had been isolated from the uninfected mice. Similar results were obtained when the cDNAs generated from the infected and control mice were tested with primers to the *C. parvum* actin gene. Negative control reactions and Southern blot verification of the amplified products were performed as described above.

DETECTION OF *C. PARVUM*

The present invention provides an improved method for diagnosis of cryptosporidiosis and further provides oligonucleotide probes and polymerase chain reaction primers specific to *C. parvum*.

5 The invention provides a method of detecting and identifying *C. parvum* specifically by identifying DNA of SEQ ID NO:1, 2, 3 or 4 or characteristic subsequences thereof by use of hybridization probing and/or nucleotide sequence amplification by the polymerase chain reaction (PCR). Amplification of these sequences by the PCR
10 provides direct indication of positive result through observation of product formation of predicted size or confirmatory hybridization probing. The specific oligonucleotide probes and primers of the invention can be applied to suitably treated samples (e.g., surface water or feces) to detect *Cryptosporidium parvum*.

15 Probing can be carried out by using radiolabeled probes in Southern Blotting, but use of other probing techniques is also provided, e.g., by tagging oligonucleotides complementary to the target sequence with a marker, such as biotin or a fluorescent agent, and relating presence of bound marker with a positive
20 result, or by using biotin tag as active agent in an ELISA in the known manner or detecting fluorescence.

The present invention particularly provides a method capable of the detection of *C. parvum* oocysts in contaminated water, a task requiring extremely sensitive techniques.

25 Typically the specific sequence amplification primers of the invention are applied to crude preparations of oocysts, fecal or

water samples, together with chain reaction reagents and the performance of the reaction is assessed under these conditions. Oocysts may be isolated by concentration from water for releasing DNA according to isolation methods well known to those of ordinary skill in the art. Amplification using PCR or Ligase Chain Reaction (LCR) can then be used to detect the presence of *Cryptosporidium*.

A first aspect of the present invention provides a method for the detection and/or identification of *C. parvum* organisms, or polynucleic acid specific thereto, comprising determining the presence of the nucleic acid sequences SEQ ID NO:1, or the complementary sequence SEQ ID NO:2, or the nucleic acid sequence SEQ ID NO:3 or its complementary sequence, SEQ ID NO:4, or characteristic subsequences of SEQ ID NO:1-4, in polynucleic acid present in or derived from a sample under investigation, and relating the presence thereof to presence of *C. parvum* or polynucleic acid thereof.

In a preferred embodiment of this aspect of the present invention the method comprises hybridization probing polynucleic acid present in or derived from a sample under investigation with an oligonucleotide that is capable of specific hybridization with sequences characteristic of the sequences SEQ ID NO:1-4 and relating the occurrence of products of the hybridization with the presence of *C. parvum* or polynucleic acid thereof. Such hybridization may be carried out by any of the conventionally known methods, eg. by Southern Blotting or one of the tagging methods referred to above.

The term "characteristic of" as used herein refers to sequences that have a sufficient number of consecutive bases which are homologous with the sequence of interest to be considered statistically highly likely to be derived from that sequence. Thus
5 herein a "characteristic sequence" is one identical to any 10 consecutive bases or more of the sequences described herein, in this case SEQ ID NO:1-4. Preferred probes for identifying such sequences are SEQ ID NO:5-8.

The specificity of the probing is controlled in the known
10 manner by selecting stringency conditions such that the chosen probe will not hybridize with sequences found in other organisms that are known to be present in or considered likely to be found in such sample. This can be determined by carrying out control experiments. Furthermore, probes can be confirmed as specific by
15 comparing their complementary sequences to sequences found in other known organisms. Such comparison can be readily made using computer databases, such as EMBL or GENBANK, whereby rapid comparison of sequences can be made.

The present invention further provides polynucleotide probes
20 suitable for this method, preferably comprising a polynucleotide subunit of one of SEQ ID NO:1-4, said probe being optionally labeled in known manner, e.g., by incorporation of a radioactive chemical or biological label (e.g., ³²P) into its structure; most preferably consisting of these sequences.

25 In a preferred embodiment the present invention provides a method for the detection and/or identification of *C. parvum*

organisms or nucleic acid specific thereto comprising use of specific sequence amplification, e.g., the polymerase chain reaction, to amplify prepared subunits of SEQ ID NO:1-4. The use of primers in the known manner may be made to target these sequences to provide an all or nothing amplification indicative of the presence of *C. parvum*.

Thus, for example, for the polymerase chain reaction, forward and reverse PCR primers having 5' end sequences SEQ ID NO:5 and SEQ ID NO:6, respectively, may be used, with or without 5' end non-hybridizing extensions:

SEQ ID NO:5 5' ATG CTA CAC AAC ATG GAG AAG G 3'

SEQ ID NO:6 5' AGT TAT CCA ATC CCA ATT GGT 3'

for example, consisting of these sequences or any other primers which specifically provide for PCR amplification of all or a portion of SEQ ID NO:1 or SEQ ID NO:2. It will be understood by those skilled in the art that when it is desired to use the ligase chain reaction rather than PCR, longer primers will be required, having a smaller gap between them on the target sequence.

In another example, preferred PCR primers found to be specific for amplification of DNA characteristic of SEQ ID NO: 3 and 4 have 5' end SEQ ID NO:7 (forward primer) and SEQ ID NO:8 (reverse primer):

SEQ ID NO:7 5' AAA ATG CCA TGT AAA TCA GCG 3'

SEQ ID NO:8 5' TCC GAA TCC AAG AAA ACC AC 3'

Specific hybridization may be carried out in each case with hybridization conditions stringent enough to exclude interfering

organisms and genomic DNA from other *Cryptosporidium* organisms while being of low enough stringency to allow hybridization with the *parvum* sequences. The determination of such conditions will be readily made by those skilled in the art by use of simple control experiments and the protocols outlined in the examples provided herein.

The present invention further provides a method for detecting or identifying *C. parvum* organisms or nucleic acid sequences specific thereto comprising:

10 (a) mixing a sample suspected of comprising said organisms and/or nucleic acid sequences with polymerase chain reaction or ligase chain reaction reagents, and respective primers targeted to specifically amplify a polynucleotide sequence within SEQ ID NO:1 and/or 2;

15 (b) subjecting the mixture to conditions under which amplification of at least one sequence characteristic of SEQ ID NO:1 and/or 2 present will occur; and

 (c) relating the production of a polymerase chain reaction product corresponding in size to the amplified subsequence to the presence of *C. parvum* organisms or nucleic acid sequences specific thereto.

20 Preferably, the polynucleotide sequence amplified is a double stranded sequence found within SEQ ID NO:1 and 2, which is amplified by PCR primers of SEQ ID NO:5 and 6 described in the sequence listing given herein, but may be any polynucleotide

25

sequence characteristic of all or a portion of SEQ ID NO:1 or SEQ ID NO:2.

The present invention further provides a method for detecting or identifying *C. parvum* organisms or nucleic acid sequences specific thereto comprising:

(a) mixing a sample suspected of comprising said organisms and/or nucleic acid sequences with polymerase chain reaction or ligase chain reaction reagents, and respective primers targeted to specifically amplify a polynucleotide sequence within SEQ ID NO:3 and/or 4;

(b) subjecting the mixture to conditions under which amplification of at least one sequence characteristic of SEQ ID NO:3 and/or 4 present will occur; and

(c) relating the production of a polymerase chain reaction product corresponding in size to the amplified subsequence to the presence of *C. parvum* organisms or nucleic acid sequences specific thereto.

Preferably, the polynucleotide sequence amplified is a double stranded sequence found within SEQ ID NO:3 and 4, which is amplified by PCR primers of SEQ ID NO:7 and 8 described in the sequence listing given herein, but may be any polynucleotide sequence characteristic of all or a portion of SEQ ID NO:3 or SEQ ID NO:4.

Again, in order to ensure specific amplification of the target sequence it is convenient to determine the degree of specificity required of the primers to be selected by either using control

experiments using DNA of organisms likely to be found in the samples to be tested as controls with which the selected primers should not initiate a polymerase chain reaction. In a further method of selecting primers, the primer nucleotide sequence contemplated for use may be compared to known sequences on 5 databases such as EMBL and Genbank where a finding of low homology with recorded sequences, particularly in the area of the primer 5' end sequence, indicates suitability for specific amplification.

Where verification of production of a PCR (or LCR) product or 10 a native unamplified sequence as being one of *C. parvum* is intended then probing may be carried out using a labeled probe specific for the sequences above, e.g., probes comprising any of the sequences SEQ ID NO:5-8 or double stranded sequences containing these. Other probe sequences selected from the aforesaid sequences will also be 15 derivable by the person skilled in the art by use of control experiments and/or databases referred to above, but it is generally recognized that sequences complementary to 10 or more consecutive bases of the target polynucleic acid will be specific enough in binding for verification of presence of such sequences.

20 Preferably, where PCR is used, the primers should be of 6 or more bases long, more preferably 10 to 30 bases long and most preferably from 15 to 21 bases long.

The unamplified or amplified polynucleotide sequence may be identified by hybridization probing using a suitably labeled 25 oligonucleotide that is capable of specific hybridization with the

sequence SEQ ID NO:1-4 or double stranded sequences made up of these, or subunits thereof.

It will be understood by those skilled in the art that when any of the methods outlined above are being applied to whole organisms, rather than to nucleic acids derived from them, it will be necessary to incorporate a procedure for liberating nucleic acids from said organisms before initiating amplification chain reaction conditions. The preferred forms of the present invention provide particular procedures whereby the sample to be tested is first suspended in a medium, e.g., a buffer, and then either sonicated or subjected to freeze/thaw cycles in buffer containing a reducing agent, e.g., dithiothreitol (DTT), in order to rupture the organisms, particularly their oocyst forms, and liberate the polynucleic acids. Particularly preferred are formats that isolate oocysts using antibodies to oocysts that have been tagged with metallic particles such that use of magnetic fields can recover individual oocyst/antibody complexes.

Using these preferred methods of the present invention it is possible to achieve detection or identification of *Cryptosporidium* organisms in oocyst form or otherwise. Preferably the sonication or freeze/thaw procedure is carried out in a medium which will form all or part of the PCR, LCR or probing medium, eg. the 1xTE medium referred to below, together with a reducing agent, eg. 1-2% DTT in buffer. Sonication is conveniently carried out using about 11 μ m peak to peak. Alternatively, for freeze/thaw procedure, the sample containers can be conveniently immersed in a cryogenic liquid such

as liquid nitrogen. Other methods for disrupting cell or oocyte structures to liberate nucleic acids into the PCR or LCR probing medium will be known to the skilled person and may be expected to be applicable before initiating the PCR, LCR and/or probing procedures.

In a still further aspect of the present invention there are provided test kits comprising one or more probes and/or primers of the present invention as described herein above, in addition to these probes and primers per se.

The method, primers and probes of the present invention will now be described by way of illustration only by reference to the following protocols and examples. Further variants of the method, primers and probes falling within the scope of the invention will occur to the person skilled in the art in the light of these.

EXAMPLE 1. POLYMERASE CHAIN REACTION DETECTION
OF C. PARVUM NUCLEOTIDE SEQUENCES

Primer pair SEQ ID NO:5 and 6 or primer pair SEQ ID NO:7 and 8 may be used in the following reaction mix/conditions to amplify a DNA fragment from *C. parvum*.

Reaction mix (volume 50 μ l)

dntp,s 250 μ moles

primers 100 pmoles of each primer (SEQ ID NO:5 and 6
or SEQ ID NO:7 and 8)

glycerol 5% v/v

Taq
polymerase 2.5 units

1XTE (10mm Tris.cl pH 8; 1mm EDTA pH8)

Buffer 5.5 mm MgCl₂
 50mm KCl
 10mm Tris.cl pH 8.3

5 plus the target nucleic acid to be amplified in a volume of
1-10 µl.

Reaction Conditions

Initial denaturing step	94°C for 3 minutes
Denature	94°C for 90 seconds
Anneal	50°C for 120 seconds 40 cycles
10 Extend	72°C for 120 seconds
Final extension	72°C for 5 minutes

15 The sample is then run on a 2% horizontal agarose gel, stained
with ethidium bromide and viewed under UV illumination and compared
to the sample gel with those from positive and negative DNA
controls.

EXAMPLE 2. PCR HYBRIDIZATION PROBING

20 The DNA on the stained gel produced in Example 1 may be
transferred onto a nylon filter (trademark-Hybond N) by Southern
Blotting, denatured and hybridized with a ³²P radiolabeled
(multiprime labeling procedure) cloned *C. parvum* internal probe
having SEQ ID NO:5 or another sequence characteristic of
SEQ ID NO:1. After washing off non-specifically bound probe, the
filter is exposed to X-ray film (autoradiography) for 12 hours or

7 days. Detection limits are respectively: 12 hours 0.3 fg, 7 days 0.03 fg of *C. parvum* DNA. It will be realized that the hybridization probe might be used directly on the sample DNA without PCR but that this will necessarily reduce sensitivity.

5 EXAMPLE 3. PREPARATION OF WHOLE OOCYSTS FOR PCR

A rapid protocol may be used for preparation of oocysts (the infective stage of *Cryptosporidium*) for PCR to avoid the long procedure associated with preparation of DNA.

1. The sample to be tested is resuspended in a solution of 1X
10 TE (see above); 1% dithiothreitol (approx. final volume is 100 μ l).
2. The oocysts are broken open by (a) 3 cycles of sonication for 10
seconds (11 μ m peak to peak) with a 50 second rest, or (b) 3
freeze/thaw cycles in liquid nitrogen. 3. The sample is then
heated to 90°C for 20 minutes. 4. Debris is pelleted by
15 centrifugation at 13000g for 5 minutes. 5. 1-10 μ l of the
supernatant is used as the template for PCR.

EXAMPLE 4. APPLICATION OF THE METHOD TO HUMAN
AND ANIMAL FECAL SAMPLES

A human fecal sample is diluted 1/100 in 1xTE/2% DTT buffer,
20 freeze thawed in liquid nitrogen five times, heated to 90°C for 20
minutes, debris pelleted at 13,000g for 5 minutes and the
supernatant used as a template for PCR, as described above, for
example, using primers described above.

UTILITY AS VACCINES

Applicants anticipate that proteins coded for by the DNA sequence described herein, in particular SEQ ID NO:1 and 3 or characteristic subsequences thereof, such as the first and second open reading frames of SEQ ID NO:1 identified above, can be used to generate antibodies or cell mediated immunity which can inhibit the pathogenesis of *C. parvum* in humans and livestock animals such as cattle. A protein such as that shown in SEQ ID NO:9 could be used in a vaccine formulation to induce production of antibodies *in vivo* for warding off infection by *C. parvum* or for the production of antibodies for use in a serum formulation against an active *C. parvum* infection.

In vivo Immunization of HumansPreparation of Antigens for Injection

Antigens and sera may be prepared for injection into humans and livestock by suitable methods well known to those of ordinary skill in the art. For example, see E.A. Kabat, Experimental Immunochimistry, 2nd edition (Thomas 1971), incorporated herein by reference. Briefly, soluble antigens which may be produced by induced plasmids containing *hemA* and 38G are generally sterilized by passage through a Chamberland, Berkefeld or Seitz filter having a pore size of 2 microns or less (appropriate filters are also available from Costar, Cambridge, MA 02140). Suspensions of insoluble antigens may be sterilized by treatment with 0.2% formalin for several days in the cold or with 0.3 to 0.5% phenol.

The antigen is then centrifuged off, washed once with sterile saline and suspended in saline. Phenol need not be removed. As a preservative for soluble as well as insoluble antigens 1% by volume of a 1% solution of methiolate is added.

5 For injection of humans with blood group substances, dextrans and pneumococcal polysaccharides, saline solutions of the desired concentration are prepared in 0.25% phenol; dextrans and the blood group substances may be autoclaved if desired. Solutions of 1 mg per ml for the dextrans and blood group substances, and 0.05 mg per
10 ml of the pneumococcal polysaccharides have been used. Sterility tests are carried out by streaking blood plates with 0.1 ml of solution and by inoculating samples into tubes containing 20 ml Difco thioglycollate broth. Plates and tubes are observed for ten days; no growth should occur.

15 Soluble proteins may be precipitated with alum to obtain an enhanced antibody response. To 100 ml sterile solution of the protein containing 1.5 mg of protein per ml, is added 5 ml of 1% sterile alum. This solution is then neutralized with 10N NaOH to maximum turbidity.

20 Vaccination schedules and dosages are also well known to those of ordinary skill in the art. Exemplary of such a schedule is to vaccinate adult humans or animals in dosages ranging from 20 to 50 to 100 to 160 μ g with booster injections provided after 28 and 56 days. (See Herrington et al., "Safety and Immunogenicity in Man of
25 a Synthetic Peptide Malaria Vaccine Against *Plasmodium falciparum* Sporozoites", Nature, 328:257-259, July 16, 1987). In another

immunization protocol, humans or other mammals may be immunized three times (days 1, 22 and 43) subcutaneously with 0.1 mg of the protein dissolved in 1 ml of PBS-3M urea per dose, or with diluent alone, and with an adjuvant such as 0.1 ml of polyalphaolefine.

5

Applications of Passive Immunization

Passive immunization can be used to treat the pathologic response to *C. parvum* in a patient. For example, anti-*C. parvum* antibodies could be administered as a serum in a hospital emergency room.

10

In vitro Immunization

The technique of *in vitro* immunization of human lymphocytes is frequently employed to generate a large variety of human monoclonal antibodies, since deliberate *in vivo* priming of humans with many antigens of interest is not feasible until approval by the Food and Drug Administration has been obtained. Techniques for *in vitro* immunization of human lymphocytes are well known to those skilled in the art. See, e.g., T. Inai et al., Histochemistry (Germany), 99(5):334-362 (May 1993); A. Mulder et al., Hum. Immunol., 36(3):186-192 (Mar. 1993); H. Harada et al., Oral Pathol. Med. (Denmark), 22(4):145-152 (April 1993); N. Stauber et al., J. Immunol. Methods (Netherlands), 161(2):157-168 (May 26, 1993); and S. Venkateswaran et al., Hybridoma, 11(6):729-739 (Dec. 1992), which are incorporated herein by reference. These techniques can be used to produce antigen-reactive human monoclonal antibodies,

20

including antigen-specific IgG, and IgM human monoclonal antibodies.

Vaccination by Plasmid Injection

It is also contemplated that vaccination may be accomplished
5 via direct intramuscular or intradermal injection of recombinant
plasmids containing hemA or 38G. Humoral as well as cell-mediated
immune response have been elicited against viral and parasite
proteins in a variety of animals using plasmid injection as is well
known to those of ordinary skill in the art. Direct DNA injection
10 may also be used to induce partial to complete immunity against
pathogen challenge. A vaccination methodology using plasmid
injection is shown below.

Plasmid DNA Construction

A cDNA such as SEQ ID NO:1 or SEQ ID NO:3 or characteristic
15 subsequences thereof of *C. parvum* is subcloned into the pRC/CMV
expression vector (Invitrogen, San Diego, CA) downstream of the
human cytomegalovirus immediate early gene 1 promoter/enhancer
region. To facilitate cloning and expression, the 5' end of the
cDNA inset is extended with a BstX I site, a mammalian ribosome
20 binding site sequence, and an ATG translational start site. The 3'
end is extended to include a Not I site. Extension is performed by
PCR using synthetic oligomer primers containing the respective
extension sequences and either the first 35 bases of the 5' end of
the cDNA or 35 bases in the distal 3' region of the cDNA. The

resulting PCR product is directionally cloned as a BstXI-NotI fragment into pRC/CMV resulting in the production of the plasmid. Duplication of the cDNA sequence and the open reading frame is confirmed by sequencing the entire insert. Plasmid DNA is prepared
5 by culture in *E. coli* strain DH5 in super broth media containing ampicillin. Following alkaline lysis, plasmid was purified by polyethylene glycol precipitation technique followed by phenol, phenol/chloroform, and two chloroform extractions.

DNA Injection and Vaccination

10 Plasmid DNA may be administered in a volume of 500 ul 0.15 M NaCl using a needle and syringe, or a jet-injection apparatus. DNA may be administered through the skin using a vaccination schedule similar to the above.

15 Changes may be made in the construction and the operation of the various components, elements and assemblies described herein or in the steps or the sequence of steps of the methods described herein without departing from the spirit and scope of the invention as defined in the following claims.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Steele, Marilyn I.; Kuhls, Thomas L.; and Nida, S. Kay
 - (ii) TITLE OF INVENTION: Detection of Cryptosporidium Parvum
 - (iii) NUMBER OF SEQUENCES: 9
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Christopher W. Corbett, Ph.D.
Dunlap & Coddling, P.C.
 - (B) STREET: 9400 North Broadway, Suite 420
 - (C) CITY: Oklahoma City
 - (D) STATE: Oklahoma
 - (E) COUNTRY: United States of America
 - (F) ZIP: 73114
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: 3.5, 1.44 Mb High Density Diskette
 - (B) COMPUTER: IBM AT Compatible
 - (C) OPERATING SYSTEM: MS-DOS 5.00
 - (D) SOFTWARE: WordPerfect 5.1 (saved in ASCII format)
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: Not Yet Assigned
 - (B) FILING DATE: Filed Herewith
 - (C) CLASSIFICATION:
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: N/A
 - (B) FILING DATE:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Christopher W. Corbett, Ph.D.
 - (B) REGISTRATION NUMBER: 36,109
 - (C) REFERENCE/DOCKET NUMBER: 5820.316
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 405-478-5344
 - (B) TELEFAX: 405-478-5349

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1298 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: no

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cryptosporidium parvum*
- (G) CELL TYPE: Unicellular Organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: INV38G

(x) PUBLICATION INFORMATION: unpublished

(K) RELEVANT RESIDUES: from 1 to 1298

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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atcctcttttc cacaaagtat ataaataagt acctgggtatc agaccataat gaaacacttt  60
ctttgcatat ctgtaccaa ataggataaa tttatgggtat atctgcgtaa tttaatagaa 120
aataaatttt tcatgagtaa taaggagtta gtagtttatt tcttacctta gaaatgccac 180
ttttaattat gttcaaatat atttgattca atttgagaat cttatcatat aaaagtttgg 240
agtatacttg tgagtatctc attatttcaa tttttcaaag tataatagaa atgcaatatt 300
aattattatt cagaagtttt ctctgtttta ttttatttta tttttcaaag tattaggaat 360
ttaggaaata attataactc cattcgggcgc caacggggccc ccacagtcac aattgacagc 420
caatgtgtga aaaaatatta attaaaagtg gaatcagaga aataatttgc atcaatctaa 480
aaagatattt ttctaaaatg tccaatatatt cagcaattaa tttcttttca gaaagtaagt 540
tttttgatgat tcttttgatga aatatatatt tctttgagtg gatttacaag taataactaa 600
cttttctatg attctataga cattggggatt actggatttg actcagaaaa tgctcctttt 660
atggcctttca aggtaagtta aataagattt aatttgcaat gtagctttta gtttaatttg 720
gattgatttt ttttatttag gaactttttg ataactctat agatgcatgt aataataagg 780

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caaaagcaga gtggcatgat tttccaaaaa aaatagaaat ctctgtgaat tttgagaatt 840
tagaattgga tgatcaggta agtaaaatac aggactagta ttaataaatt tgatttttta 900
ttaaattgg tttgaacttt aaggcaagaa taaacatcat tgtaagagat acaggatgcg 960
gaataccttt agaatccatt gacctattgg gaactctttt tggaacaaag tgagttaa 1020
tattaattgt atttaaagt attgattaaa tttttaattc tgattcatct tttagtaaga 1080
aaataaataa aagagattct tattatacag gacaatttgg ggtaggcctt aaaatgatat 1140
tactttatgc tacacaacat ggagaaggaa acgttaaagt taagatgaga atgggaaaca 1200
agatttggga gtttactctg ctttgtaacc tgaatgatgg atctttttat gtagggaata 1260
gtgagagttt tgattatacc aattgggatt ggataact 1298

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1298 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTISENSE: yes

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cryptosporidium parvum*
- (G) CELL TYPE: Unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: INV38G

(x) PUBLICATION INFORMATION: unpublished

(K) Relevant residues: from 1 to 1298

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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taggagaaag gtgtttcata tatttattca tggaccatag tctggtatta ctttgtgaaa   60
gaaacgtata gacatgggtt tctctatctt aaataccata tagacgcatt aaattatctt  120
ttatttaaaa agtactcatt attcctcaat catcaaataa agaatggaat ctttacggtg   180
aaaattaata caagtttata taaactaagt taaactctta gaatagtata ttttcaaacc   240
tcatatgaac actcatagag taataaagtt ataaagtttc atattatctt tacgttataa   300
ttaataataa gtcttcaaaa gagacaaaat aaaataaaat aaaaagtttc ataactccta   360
aatcctttat taatattgag gtaagccgcg gttgcccggt ggtgtcagta ttaactgtcg   420
gttacacact tttttataat taattttcac cttagtctct ttattaaacg tagttagatt   480
tttctataaa aagattttac aggttataaa gtcgttaatt aaagaaaagt ctttcattca   540
aaaaacacta agaaaacact ttatatataa agaaactcac ctaaagtgtc attattgatt   600
gaaaagatac taagatatct gtaaccctaa tgacctaaac tgagtctttt acgaggaaaa   660
taccgaaagt tccattcaat ttattctaaa ttaaactgta catcgaaaat caaattaaac   720
ctaactaaaa aaaataaatc cttgaaaaac tattgagata tctacgtaca ttattattcc   780

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gttttcgtct caccgtacta aaagggtttt tttatcttta gagacactta aaactcttaa 840
atcttaacct actagtccat tcattttatg tcctgatcat aattatttaa actaaaaaat 900
aattttaacc aaacttgaaa ttccgttctt atttgtagta acattctcta tgccttacgc 960
cttatggaaa tcttaggtaa ctggataacc cttgagaaaa accttgtttc actcaaattt 1020
ataattaaca taaatttcac taactaattt aaaaattaag actaagtaga aaatcattct 1080
tttatttatt ttctctaaga ataatatgtc ctgttaaacc ccatccggaa ttttactata 1140
atgaaatacg atgtgttgta cctcttcctt tgcaatttca attctactct taccctttgt 1200
tctaaaccct caaatgagac gaaacattgg acttactacc tagaaaaata catcccttat 1260
cactctcaaa actaatatgg ttaaccctaa cctattga 1298

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 943 Base Pairs
- (B) TYPE: Nucleic Acid
- (C) STRANDEDNESS: Single
- (D) TOPOLOGY: Linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: no

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cryptosporidium parvum*
- (G) CELL TYPE: unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: Hema

(x) PUBLICATION INFORMATION: unpublished

- (K) RELEVANT RESIDUES: from 1 to 943

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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aatatactat aaattaatcc tatattttta atttataatt gaaactaatt aaataaaata      60
ataaggtgaa atg agc caa aaa ata gat ttg aat caa ggt gaa aga act gag    112
      Met Ser Gln Lys Ile Asp Leu Asn Gln Gly Glu Arg Thr Glu
      1              5              10

tca aaa tgt cca gaa acg ctt aat gaa ttt tct aat atg tat aag cat aaa    163
Ser Lys Cys Pro Glu Thr Leu Asn Glu Phe Ser Asn Met Tyr Lys His Lys
15              20              25              30

aaa atg cca tgt aaa tca gcg ctg aac ttt ggt gaa aat att ata aat atg    214
Lys Met Pro Cys Lys Ser Ala Leu Asn Phe Gly Glu Asn Ile Ile Asn Met
      35              40              45

gga gct cca agt ttt ata cca gaa tat cca aca gct tat tat tct aac cca    265
Gly Ala Pro Ser Phe Ile Pro Glu Tyr Pro Thr Ala Tyr Tyr Ser Asn Pro
      50              55              60              65

gaa gtt atg aat caa gga ata aat tca tta gtg gat aca aaa caa cta tat    316
Glu Val Met Asn Gln Gly Ile Asn Ser Leu Val Asp Thr Lys Gln Leu Tyr
      70              75              80

gac tat tca tcc cct gta ggt gat ctt gaa aaa acc att gaa cat tac aaa    367
Asp Tyr Ser Ser Pro Val Gly Asp Leu Glu Lys Thr Ile Glu His Tyr Lys
      85              90              95

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atg agt cat gaa ata gga tgg aat gca agt aat tct ttc act cct aca aat	418
Met Ser His Glu Ile Gly Trp Asn Ala Ser Asn Ser Phe Thr Pro Thr Asn	
100 105 110 115	
tct ggt tct ttg gaa ctt ttc caa ttt gaa aga aga gat agt cct gtc gct	469
Ser Gly Ser Leu Glu Leu Phe Gln Phe Glu Arg Arg Asp Ser Pro Val Ala	
120 125 130	
gtt gat act tta aaa aat tac cca tct ttt gag aga atg aat agt ggt ttt	520
Val Asp Thr Leu Lys Asn Tyr Pro Ser Phe Glu Arg Met Asn Ser Gly Phe	
135 140 145 150	
ctt gga ttc gga aga aaa cct tct aat tca ata ttt tct att gga caa aaa	571
Leu Gly Phe Gly Arg Lys Pro Ser Asn Ser Ile Phe Ser Ile Gly Gln Lys	
155 160 165	
ctt gaa aga ctc tct tct aat gaa ata atc aac aat atc aat acc ctt cct	622
Leu Glu Arg Leu Ser Ser Asn Glu Ile Ile Asn Asn Ile Asn Thr Leu Pro	
170 175 180	
gaa att gaa aat ggt att gag aac ggt att gaa aat gga att gaa aat gga	673
Glu Ile Glu Asn Gly Ile Glu Asn Gly Ile Glu Asn Gly Ile Glu Asn Gly	
185 190 195 200	
gat att cct cac gtt gat ttg gag caa tat aca caa tta tct tac ttt gaa	724
Asp Ile Pro His Val Asp Leu Glu Gln Tyr Thr Gln Leu Ser Tyr Phe Glu	
205 210 215	
aaa tat atc aat ttt aac att gac caa att aag tgaatccatt tcttaaatat	777
Lys Tyr Ile Asn Phe Asn Ile Asp Gln Ile Lys	
220 225	
aactactttt tataacccaaa aatgctatga aacaaatcat ttttgcaaaa tatattggaa	837
tattattaga tttatgaaat atcaagctgt tcaaaagaat agcctgatac tccacgtaaa	897
atttgtaata ttgaaaagga atagtaactc ccaggaataa tacaaa	943

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 943 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTISENSE: yes

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cryptosporidium parvum*
- (G) CELL TYPE: Unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: Hema

(x) PUBLICATION INFORMATION: unpublished

- (K) Relevant residues: from 1 to 943

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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ttatatgata ttttaattagg atataaaaat taaatattaa ctttgattaa tttattttat 60
tattccactt tactcggttt tttatctaaa cttagttcca ctttcttgac tcagttttac 120
aggtctttgc gaattactta aaagattata catattcgta tttttttacg gtacatttag 180
tcgcgacttg aaaccacttt tataatatatt ataccctcga gggtcaaaat atgggtcttat 240
aggttgtcga ataataagat tgggtcttca atacttagtt ccttatttaa gtaatcacct 300
atgttttggt gatatactga taagtagggg acatccacta gaactttttt ggtaacttgt 360
aatgttttac tcagtacttt atcctacctt acgttcatta agaaagtgag gatgtttaag 420
accaagaaac cttgaaaagg ttaaactttc ttctctatca ggacagcgac aactatgaaa 480
ttttttaatg ggtagaaaac tctcttactt atcaccaaaa gaacctaagc cttcttttgg 540
aagattaagt tataaaagat aacctgtttt tgaactttct gagagaagat tactttatta 600
gttggtatag ttatgggaag gactttaact ttaccataa ctcttgccat aacttttacc 660
ttaactttta cctctataag gagtgcact aaacctcgtt atatgtgtta atagaatgaa 720
actttttata tagttaaaat tgtaactggt ttaattcact taggtaaaga atttatattg 780

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atgaaaaata ttggttttta cgatactttg tttagtaaaa acgttttata taaccttata 840
ataatctaaa tactttatag ttcgacaagt tttcttatcg gactatgagg tgcattttta 900
acattataac ttttccttat cattgagggg ccttattatg ttt 943

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: yes

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Cryptosporidium parvum
- (G) CELL TYPE: Unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: INV38G

(x) PUBLICATION INFORMATION: unpublished

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATG CTA CAC AAC ATG GAG AAG G

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: yes

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Cryptosporidium parvum
- (G) CELL TYPE: Unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: INV38G

(x) PUBLICATION INFORMATION: unpublished

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

AGT TAT CCA ATC CCA ATT GGT

21

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: no

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cryptosporidium parvum*
- (G) CELL TYPE: Unicellular organism

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: HemA

(x) PUBLICATION INFORMATION: unpublished

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AAA ATG CCA TGT AAA TCA GCG

21

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: genomic DNA

(iii) HYPOTHETICAL: no

(iv) ANTI-SENSE: yes

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Cryptosporidium parvum
- (G) CELL TYPE: Unicellular

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: genomic
- (B) CLONE: Hema

(x) PUBLICATION INFORMATION: unpublished

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TCC GAA TCC AAG AAA ACC AC

20

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 229 Amino acids
 (B) TYPE: Amino acid
 (D) TOPOLOGY: Linear

(ii) MOLECULAR TYPE: protein

(x) PUBLICATION INFORMATION: unpublished

(K) RELEVANT RESIDUES: from 1 to 229

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

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Met Ser Gln Lys Ile Asp Leu Asn Gln Gly Glu Arg Thr Glu Ser Lys
1           5           10           15
Cys Pro Glu Thr Leu Asn Glu Phe Ser Asn Met Tyr Lys His Lys Lys
          20           25           30
Met Pro Cys Lys Ser Ala Leu Asn Phe Gly Glu Asn Ile Ile Asn Met
          35           40           45
Gly Ala Pro Ser Phe Ile Pro Glu Tyr Pro Thr Ala Tyr Tyr Ser Asn
          50           55           60
Pro Glu Val Met Asn Gln Gly Ile Asn Ser Leu Val Asp Thr Lys Gln
65           70           75           80
Leu Tyr Asp Tyr Ser Ser Pro Val Gly Asp Leu Glu Lys Thr Ile Glu
          85           90           95
His Tyr Lys Met Ser His Glu Ile Gly Trp Asn Ala Ser Asn Ser Phe
          100          105          110
Thr Pro Thr Asn Ser Gly Ser Leu Glu Leu Phe Gln Phe Glu Arg Arg
          115          120          125
Asp Ser Pro Val Ala Val Asp Thr Leu Lys Asn Tyr Pro Ser Phe Glu
          130          135          140
Arg Met Asn Ser Gly Phe Leu Gly Phe Gly Arg Lys Pro Ser Asn Ser
145          150          155          160
Ile Phe Ser Ile Gly Gln Lys Leu Glu Arg Leu Ser Ser Asn Glu Ile
          165          170          175
Ile Asn Asn Ile Asn Thr Leu Pro Glu Ile Glu Asn Gly Ile Glu Asn
          180          185          190
Gly Ile Glu Asn Gly Ile Glu Asn Gly Glu Ile Pro His Val Asp Leu
          195          200          205

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Glu Gln Tyr Thr Gln Leu Ser Tyr Phe Glu Lys Tyr Ile Asn Phe Asn
210 215 220

Ile Asp Gln Ile Lys
225

What is claimed is:

1. A method for the detection of *Cryptosporidium parvum* organisms, or nucleic sequences specific thereto, comprising determining the presence of a nucleic acid sequence characteristic of at least one of the nucleic acid sequences SEQ ID NO:1, SEQ ID
5 NO:2, SEQ ID NO:3, and SEQ ID NO:4, in nucleic acid present in or derived from a sample under investigation, and relating the presence thereof to the presence of *Cryptosporidium parvum* or nucleic acid thereof.

2. The method of claim 1 wherein the method further comprises using a specific sequence amplification technique which requires an oligonucleotide primer that is capable of hybridization with a sequence characteristic of at least one of SEQ ID NO:1 and
5 its complementary sequence SEQ ID NO:2 thereby forming an amplification product and relating the production of the amplification product to the presence of *Cryptosporidium parvum* or said nucleic acid.

3. The method of claim 2 wherein in the step of using a specific sequence amplification technique, the oligonucleotide primer comprises from 10 to 30 consecutive bases characteristic of at least one of nucleic acid sequences SEQ ID NO:1 and SEQ ID NO:2.

4. The method of claim 3 wherein the oligonucleotide primer further comprises from 12 to 25 consecutive bases.

5. The method of claim 4 wherein the oligonucleotide primer further comprises from 15 to 22 consecutive bases.

6. The method of claim 2 wherein the step of using a specific sequence amplification technique is carried out using the polymerase chain reaction and wherein the reaction uses a forward primer and a reverse primer.

7. The method of claim 6 wherein the forward and reverse primers have sequences SEQ ID NO:5 and SEQ ID NO:6, respectively.

8. The method of claim 1 wherein the method further comprises using a specific sequence amplification technique which requires an oligonucleotide primer that is capable of hybridization with a sequence characteristic of at least one of SEQ ID NO:3 and
5 its complementary sequence SEQ ID NO:4 thereby forming an amplification product and relating the production of the amplification product to the presence of *Cryptosporidium parvum* or said nucleic acid.

9. The method of claim 8 wherein in the step of using a specific sequence amplification technique, the oligonucleotide primer comprises from 10 to 30 consecutive bases characteristic of at least one of nucleic acid sequences SEQ ID NO:3 and SEQ ID NO:4.

10. The method of claim 9 wherein the oligonucleotide primer further comprises from 12 to 25 consecutive bases.

11. The method of claim 10 wherein the oligonucleotide primer further comprises from 15 to 22 consecutive bases.

12. The method of claim 8 wherein the step of using a specific sequence amplification technique is carried out using the polymerase chain reaction and wherein the reaction uses a forward primer and a reverse primer.

13. The method of claim 12 wherein the forward and reverse primers have sequences SEQ ID NO:5 and SEQ ID NO:6, respectively.

14. A method for the detection of *Cryptosporidium parvum* organisms or nucleic acid sequences specific thereto comprising:

5 (a) mixing a sample suspected of comprising said organisms and/or nucleic acid sequences thereof in a specific sequence amplification reaction using specific sequence amplification reaction reagents and primers for said amplification, said primers being targeted to specifically amplify a polynucleotide sequence characteristic of at least one of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4;

10 (b) subjecting the mixture to conditions under which the amplification of any of said sequences will occur, thereby forming an amplification product; and

(c) relating the production of any amplification product to the presence of *Cryptosporidium parvum* or nucleic acid sequences specific thereto.

15
5 15. The method of claim 14 wherein the specific sequence amplification reaction is the polymerase chain reaction and the primers used have sequences SEQ ID NO:5 and SEQ ID NO:6 for amplifying a sequence characteristic of at least one of SEQ ID NO:1 and SEQ ID NO:2.

16. The method of claim 14 wherein the specific sequence amplification reaction is the polymerase chain reaction and the primers used have sequences SEQ ID NO:7 and SEQ ID NO:8 for amplifying a sequence characteristic of at least one of SEQ ID NO:3 and SEQ ID NO:4.

17. The method of claim 14 further comprising liberating nucleic acids from said organism before initiating amplification chain reaction conditions.

18. The method of claim 17 wherein the liberating step further comprises either sonicating or subjecting the organisms to freeze/thaw cycles in order to rupture the organisms and liberate the nucleic acids therefrom.

19. The method of claim 14 wherein the mixing step is preceded by first suspending the organisms in a medium.

20. The method of claim 14 wherein in the mixing step the primers are labeled by incorporation of a radioactive, chemical or biological label into their structure.

21. A diagnostic kit comprising at least one primer suitable for the amplification of a sequence characteristic of at least one of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4 for detecting *Cryptosporidium parvum* in a sample.

22. The diagnostic kit of claim 21 wherein the primer is further defined as being labeled by incorporation of a radioactive, chemical or biological label into its structure.

23. A DNA of sequence *Cryptosporidium parvum* comprising at least one of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4.

24. The DNA sequence of claim 23 incorporated into a vector.

25. A polypeptide coded for by one of the nucleic acid sequences SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4, or by a subsequence characteristic thereof.

26. An oligonucleotide primer comprising from 10 to 30 consecutive bases characteristic of at least one of nucleic acid sequences SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4.

27. An antibody produced in response to a polypeptide or peptide encoded by at least one of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4 and characteristic subsequences thereof.

28. A composition for inducing an antibody response to *Cryptosporidium parvum* when administered in an effective dose to a mammal in a pharmaceutically acceptable vehicle for administration to the mammal wherein the composition comprises a polypeptide or peptide encoded by at least one of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4 and characteristic subsequences thereof.

29. The composition of claim 28 further comprising an adjuvant.

30. A plasmid comprising a DNA sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 and SEQ ID NO:4 or characteristic subsequences thereof.